Interactions of physico-chemical properties of the River Nile water and fungal diversity in River Nile streams in Delta region

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Abstract

Water pollution in Egypt is increasing over time, with constant supply of pollutant effluents to the water system from several sources. Water samples were collected for hydrochemical analysis during four successive seasons from some irrigation canals and streams of River Nile in Delta region governorates. There are various correlations patterns between fungal communities and physico-chemical parameters. Air and water temperature were major factors that affect fungal distribution. Calcium and magnesium have inverse correlation on fungal diversity. There was no significant correlation of SO_4^{2-} and NO_2^{-} with distribution of freshwater fungi. Ammonia and phosphate encouraged fungal growth. Salinity played active role in determining fungal diversity and community composition. Changes in species composition were correlated positively with elevated concentrations of NH_3^+ , and pH in some governorates.

Key word: Physico-chemical, populations, Nile delta, Freshwater fungi.

Introduction:

Fungi may be exposed to a wide variety of organic and inorganic pollutants in the environment. Pollutants may exhibit toxicity and cause changes in fungal community composition (Gadd 2007). The effect of pollutants on fungal population community size and composition is particularly difficult to assess. Environmental pollution might be expected to lead to both toxic (destructive) and enrichment disturbance on fungal populations (Wainwright 1988). The resultant degree of toxic disturbance will depend upon both toxicant concentration and its availability to the fungal population, as well as to the susceptibility of the individuals involved. Toxicants may be have a selectively effect on a few fungal species, or amore generalized effect.

Freshwater fungi are considered a cosmopolitan and a diverse group

(McLaughlin et al., 2001). Lignicolous freshwater fungi grow on submerged woody debris in freshwater streams, ponds and lakes (Goh and Hyde 1996, Wong et al., 1998). Lignicolous freshwater fungi play important role in the decomposition of by breaking submerged wood down lignocelluloses and releasing nutrients which are important in ecosystem functioning (Yuen et al., 1998, Bucher et al., 2004, Hyde et al., 2016).

Several studies reported the effect of pollutants on freshwater fungal communities (Tan and Lim 1984; Maltby and Booth 1991; Au *et al.*, 1992; Bärlocher 1992a; Tsui *et al.*, 2001; Luo *et al.*, 2004). The fungal community structure greatly differs with the physico-chemical properties of the respective habitats, such as water flow (Pattee and Chergui 1995; Baldy *et al.*, 2002), nutrient concentrations (Gulis *et al.*, 2004; Rankovic

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2005), salinity (Hyde and Lee1995; Roache et al., 2006), temperature (Bärlocher et al., 2008; Raja et al., 2009) and depth (Wurzbacher et al., 2010). The ability of fungi to survive in the presence of potentially toxic metals depends on a number of biochemical and structural properties, including physiological and/or genetically adaptation (Gadd 2000, 2007). In general terms, toxic metals may affect fungal populations by reducing abundance and species diversity (Babich and Stotzky 1985; Arnebrant et al., 1987).

Water pollution in Egypt is increasing over time, with constant supply of pollutant effluents to the water system from several sources. One of the major sources is the disposal of untreated or semi-treated domestic wastes into water bodies. The low level of sanitation service especially in rural areas makes nearby streams (either canals or drains) the perfect places for inhabitants to dispose their sewage. Moreover, the excessive use of fertilizers and pesticides is another major source of water pollution. The effect of agricultural pollution was increased due to the extensive drainage reuse within the Nile Delta. Several studies revealed that untreated industrial wastes of more than 350 factories were discharged directly into the Nile and the Mediterranean Sea, most of them released known toxic and hazardous explicitly chemicals such as detergents, heavy metals and pesticides (RNPD 1989).

Materials and Methods:

Water samples were collected in clean oneliter polyethylene bottles for hydrochemical analysis during four successive seasons from irrigation canals and the River Nile in Delta region, during the period between February 2010 and December 2011.

Submerged, dead woody debris were collected randomly from irrigation canals and the River Nile, four times over 2 years. Sampling times included the four seasons (winter, summer, spring, and autumn) in the eight governorates (Dakahlya, Sharqiyah,

Qalyubiya, Kafr El-Sheikh, El-Behera, Gharbeyia, Menofya and Damietta) within the Nile Delta region.

Chemical analysis of the collected water samples was carried out according to the standard methods (Rainwater *et al.*, 1960; Booth 1983). Fungal species richness, number of fungal collections per sample and number of records for each taxonomic group in the studied governorate and season collection were calculated. Canonical Correlation Analysis (CCA) was employed to test if variation in community composition is explained by physicho-chemical parameters.

Hydrochemical analysis:

Air and water temperature at each site were measured *in situ* using Celsius-thermometer. Potential of Hydrogen (pH) values of water samples were measured using a digital pH meter (HANNA pH 211 microprocessor pH meters).

Electrical conductivity (EC μ s/cm) of water samples was measured using EC meter (model: HANNA HI 99300 Conductivity meter). Salinity, total dissolved solids (TDS) in mg/l were measured for each sample using (HANNA HI 99300 Conductivity meter). Alkalinity was determined by titration using phenolphthalein and methyl orange indicators.

Chloride $(C1^{-})$ anion content determined by volumetrically titration against silver nitrate using potassium chromate (K₂CrO₄) as an indicator. Hardness was determined by titration of Erichrome black T indicator and standard (0.01)Ethylenediaminetetraacetic acid (EDTA) solution. Calcium (Ca 2+) and Magnesium (Mg ²⁺) cations content of water samples were determined by complexometric titration using EDTA solution and Eriochrome black T as indicator. Calcium was determined using muroxide indicator in presence of sodium hydroxide, while magnesium cation was calculated by subtracting the calcium value from (Ca 2+ and Mg 2+) value after their determination using Eriochrome black T.

Sulphate (SO₄²-) was determined in water samples by the turbidimetric method. It is based upon the fact that barium sulphate tends to precipitate in a colloidal form. The absorbance of the barium sulphate solution is measured by turbidimeter and the sulphate ion concentration, determined by comparison of the reading with a standard curve. Ammonia (NH₃) concentration was determined in water samples using phenate method. Ammonia reacts with alkaline phenol and hypochlorite to form indophenol blue that is proportional to the ammonia concentration. The blue color is intensified with sodium nitroprusside to form a colored complex that absorbs at 660 nm. Concentration of ammonia is determined by comparison of absorbance signal with calibration results obtained from prepared standards of varying concentrations (APHA 2005).

Phosohate (PO₄³⁻) anion concentration was determined colorimetrically using UV/visible spectrophotometer, at wavelength 700 nm, using molybdate reagent and Tin chloride (SnCl₂) as indicator.

Nitrite (NO₂⁻) ion was determined by reaction of nitrite ions with sulfanylamide in acidic medium and the diazo compound obtained further reacts with diamine yielding in an azo color. The nitrite ion concentration is determined by measuring the absorbance of the azo color at 540-545 nm by spectrophotometer.

Statistical analysis: Variation in averages of physico-chemical parameters and fungi distribution among tested governorates was analysed using PAST software version 2.11 (Hammer *et al.*, 2001). Canonical correspondence analysis (CCA) included data of 218 fungal taxa and 11 environmental parameters across 8 sampling governorates during 4 season's period was applied to evaluate role of environmental variables in fungal communities.

Results:

Physical and chemical parameters varied between sites, governorates and seasons. Interaction between physico-chemical properties and diversity of freshwater fungi was strongly observed. The minimum, maximum and average values of hydrochemical analysis of the collected water samples were listed in table 1. The patterns of their variations along the studied area and during study period are illustrated in figs. 1 to 7.

Physico-chemical characteristics of River Nile water:

The air temperature ranged between 16 and 44 °C during the studied period. The maximum air temperature, 44 °C, was recorded in August 2010 while the minimum temperature was recorded in December 2011 (Table 1). The water temperature ranged from 10 to 29 °C. The highest water temperature was 29 °C in August 2010 while the lowest was 10 °C in December 2011 (Table 1). Potential of Hydrogen (pH) values ranged from 6 to 6.8 in the present study (Table 2).

Total dissolved salts (TDS) content of water samples were fluctuated between 227 ppm and 614 ppm. The highest values of TDS were reported in Kafr El-Sheikh (375), Damietta (328)and Sharqiyah (313)governorates, whereas the lowest ones Gharbeyia occurred and Menofya governorates (Tables 1, 2 and Figure 1).

Electrical conductivity (EC) values exhibited great variations between the different governorates and seasons and were fluctuated between 324 µs/cm and 932 µs/cm in the different sites in Nile Delta region. The highest averages of EC were reported from Kafr El-Sheikh (583), Sharqiyah (529) and Menofya (502). Alkalinity of studied streams and irrigation canals in the different four seasons was ranged between 88 and 320 mg/L. Kafr El-Sheikh and El Behera governorates had the highest values of alkalinity (Table 1). Chloride concentrations ranged between 15 and 90 ppm (Table 1). The highest values of chloride concentration were reported Menofya and Qalyubiya governorates. In

August 2010 collection (28-90 ppm) while the lowest values in February 2010 (15-23 ppm). Calcium and magnesium concentrations ranged between 25 and 57.5 ppm, 13.8 and 31.8 ppm, respectively. The average values of calcium and magnesium concentrations showed narrow differences between governorates and seasons collections. Nitrite concentration in water samples was fluctuated between 0.015 and 0.15 ppm while, Ammonia concentrations ranged between 0.01 and 1.76 ppm in the studied sites as shown in Table (1). Sulphate ion concentrations ranged between

19.6 and 46 ppm. Meanwhile phosphate was in the range from 0.001 to 0.39 ppm (Table 1). The values of the physical and chemical parameters of the different seasons and governorates were listed in tables 2 and 3.

Some physico-chemical parameters exhibited strong peak in some studied governorates or seasons. For example, NH₃+and Cl⁻ concentration were very high in Menofya and Qalyubiya governorates and August collection, TDS peaked in Kafr El-Sheikh and Damietta and also Ca ²⁺ peaked in KafrEl-Sheikh.

						Fe	brua	ry 20	010 co	llectio	n						
Site		Physic	cal prop	erties			Chemical properties										
Site	A.T 0C	W.T 0C	pН	TDS ppm	EC µS/cm	Alkalinity mg/L	Cl ⁻	Ca ²⁺ ppm	Mg ²⁺ ppm	SO ₄ ² - ppm	NO ₂ - ppm	NH ₃ ppm	PO ₄ 3- ppm	S.R	NRS		
Dakahlya			7.5	307.75	479.3	161.3	16.5	39.25	17	31.49	0.024	0.33	0.22	42	0.67		
Sharqiyah	Min. temperature (8 -18) Max. emperature (16-34)		6.9	227	442	130	15	34	15.4	35.8	0.009	0.023	0.09	31	1.12		
Qalyubiya	e (8 (16		6.5	393.75	626.5	206.5	23.25	52.25	22.75	36.87	0.022	0.97	0.32	43	0.90		
Kafr El-Sheikh	ture	16-18	6.6	281	480	104	17	32	19.2	29.7	0.032	0.037	0.12	20	0.90		
El Behera	pera	16	6.7	408.5	650	247	22.5	52	22	40.5	0.041	0.46	0.06	23	0.82		
Gharbeyia	tem)		7.4	258	411.5	126.5	14.5	33	18	30.5	0.0015	0.045	0.015	30	0.73		
Menofya	lin. Tax		6.9	280	446	88	15	36	27	32	0.003	0.6	0.11	24	1.24		
Damietta	W		6.8	310	498	112	19	40	22	33	0.044	0.04	0.07	20	0.95		
Average			6.9	308.25	504.16	146.91	17.84	39.81	20.42	33.73	0.02	0.31	0.13	29	0.92		
Range			6. 5 - 7.5	227-408	411-650	88-247	15-23	32-52	15-27	29.7-40.5	.0015044	.023-0.97	.01532	20-43	0.67-1.24		
	August 2010 collection																
Dakahlya			6.5	255.8	388.7	192.3	23.0	37.0	21.0	25.5	0.05	1.05	0.15	58	1.07		
Sharqiyah	<u>8</u> €		6.1	463.7	703.67	247.7	45.0	50.33	27.13	35.9	0.03	0.92	0.07	37	1.02		
Qalyubiya	(32 -44)		6.6	274.7	434	185.7	55.67	45.13	20.37	31.13	0.08	1.31	0.21	39	0.83		
Kafr El-Sheikh	9 a	.59	6.3	614.5	932.0	320.0	48.0	57.5	31.8	19.85	0.08	0.88	0.39	38	1.27		
El Behera	ratu	27-29	6.3	267	406	178	38	38	19	23	0.037	1.27	0.2	24	1.22		
Gharbeyia	Min. temperature Max. temperature		6.2	237.5	360.5	137.5	31.5	29.0	19.5	35.5	0.07	1.76	0.02	44	1.12		
Menofya	lin. t		6	303	460	105	90	40	16.8	22.5	0.08	1.5	0.2	35	1.78		
Damietta	N		6.7	414	577	244	28	44	24	23	0.019	0.68	0.09	23	0.80		
Average			6.3	353.78	532.73	201.3	44.9	42.62	22.45	27.05	0.06	1.17	0.17	37	1.14		
Range			6 - 6.7	237-614	360-932	105-320	23-90	25-57.5	16.8-31.8	19.8-35.9	.01908	.68-1.76	.0239	23-58	0.8-1.78		
							May	201	l colle	ction							
Dakahlya			7.68	168.5	336.5	121	27.75	27.0	16.4	22.18	0.01	0.09	0.15	56	1.21		
Sharqiyah	£ 6		7.5	186	383	139	30	35	20	23.97	0.01	0.07	0.07	30	1.19		
Qalyubiya	(10 - 25) (22 - 40)		7.2	187.3	372.7	121.3	33.3	32.8	21.2	21.5	0.05	0.06	0.08	49	0.81		
Kafr El-Sheikh	ure	22	7.6	244	410	141	34	45	23	35.7	0.06	0.15	0.06	24	1.23		
El Behera	perat	20-22	7.5	180	360	133	29	30	17	24.4	0.025	0.025	0.09	30	1.09		
Gharbeyia	Min. temperature Max. temperature	``	7.6	175	350	110	23	25.2	17	24.4	0.02	0.01	0.015	43	1.03		
Menofya	Min. Max.		7.5	188	355	140	34	30	18	30.3	0.077	1.48	0.02	36	1.0		
Damietta			7.65	198.5	400	146.5	35	42	22.5	34.3	0.04	0.23	0.03	37	1.26		
Average			7.46	184.3	367.67	128.5	31.2	32.5	19.53	25.16	0.03	0.21	0.07	38	1.10		
Range			7.2 -7.7	168-244	336-400	110-1469	23-35	25-45	16.4-23	19.6-39.1	0.01-0.07	0.01-1.48	0.01-0.15	24-56	0.8-1.26		

				December 2011 collection											
			Physic	cal prop			Fungal population								
	A.T 0C	W.T 0C	pН	TDS ppm	EC μS/cm	Alkalinity mg/L	Cl ⁻	Ca ²⁺ ppm	Mg ²⁺ ppm	SO ₄ ²⁻ ppm	NO ₂ · ppm	NH ₃ ppm	PO ₄ 3. ppm	S.R	NRS
Dakahlya			7.7	323	505	172	49	31	15.2	39	0.06	0.26	0.012	52	0.95
Sharqiyah	17)		7.8	377	589	193	55	36	18.8	45	0.15	0.4	0.08	24	0.73
Qalyubiya	(6 - (18		7.4	315	448	173	47	33	16	37	0.008	0.2	0.02	42	1.05
Kafr El-Sheikh	ure	12	7.3	364	510	207	31	35	20	35.7	0.04	0.25	0.08	24	0.85
El Behera	temperature temperature	10-	7.4	380	460	213	39	31	19	24.4	0.02	0.035	0.06	15	0.76
Gharbeyia			7.5	300	468	178	34	33	17.5	44	0.03	0.1	0.02	43	0.91
Menofya	Min. Max.		7.5	310	746	169	45	34	17.5	46	0.04	0.3	0.013	41	0.9
Damietta			7.6	390	433	165	40	29	14.3	41	0.08	0.19	0.013	30	0.83
Average			7.53	345	520	183.8	42.5	32.8	17.3	39.01	0.05	0.2	0.04	33.8	87.3
Range			7.3-7.8	300-390	433- 746	165-213	31-55	29-36	14.3-20	24.4-46	.008-0.15	0.03-0.4	0.001-0.08	15-52	0.73-1.05

Table (1): The variation in physical and chemical parameters of the collected water samples from the eight governorates in different seasons.

A.T = Air temperature; W.T = Water temperature, TDS = Total dissolved salts: EC = Electric conductivity; S.R = Species richness; NRS = number of fungal collections per sample.

		Physica	l proper	ties			Chemi	Fungal records number								
Site	pН	TDS	EC	Alkalinity	Cl ⁻	Ca ²⁺	Mg ²⁺	SO ₄ ² -	NO ₂ -	NH ₃	PO43-	NRS	As R	An R	BR	M R
Dakahlya	7.35	263.76	427.4	161.6	29.1	33.56	17.40	29.54	0.04	0.43	0.13	0.93	396	246	17	58
Sharqiyah	7.08	313.43	529.4	177.4	36.3	38.83	20.33	35.17	0.05	0.35	0.08	1.07	182	97	11	47
Qalyubiya	6.93	292.69	470.3	171.6	39.8	40.8	20.08	31.63	0.04	0.64	0.16	0.91	264	200	10	51
Kafr El-Sheikh	6.95	375.88	583.0	193.0	32.5	42.38	23.50	30.24	0.05	0.33	0.16	1.19	144	111	5	63
El Behera	6.98	308.88	469.0	192.7	32.1	37.75	19.25	28.08	0.03	0.45	0.10	1.02	135	69	3	30
Gharbeyia	7.18	242.63	397.5	138.0	25.7	30.05	18.00	33.60	0.03	0.48	0.02	0.96	295	185	9	50
Menofya	6.98	270.25	501.7	125.5	46.0	35.00	19.83	32.70	0.05	0.97	0.09	1.2	178	144	9	17
Damietta	7.19	328.13	477.0	166.9	30.5	38.75	20.70	32.83	0.05	0.29	0.05	1.04	204	102	0	12

Table (2): The correlation between average of physico-chemical parameters and the fungal records numbers in the studied governorates.

	Physical properties								Ch	emic	al pr	oper	tiess	Fungal records number					
Seasons	Min A.T ⁰ C	Max A.T °C	W.T °C	pН	TDS ppm	EC µS/cm	Alkalinity	Cl ⁻		-			NH ₃ ppm	PO ₄ ^{3.} ppm	NRS	As R	An R	B R	M R
February 2010 (winter)	8-18	16-34	16-18	6.9	308.3	504.2	146.9	17.84	39.8	20.4	33.7	0.02	0.31	0.13	0.89	311	285	14	98
August 2010 (summer)	20-28	32-44	27-29	6.3	353.8	532.7	201.3	44.9	42.6	22.5	27.1	0.06	1.17	0.17	1.1	463	262	23	138
May 2011 (last spring)	10-25	22-40	20-22	7.46	184.3	367.7	128.5	31.21	32.5	19.5	25.2	0.03	0.21	0.07	1.05	552	280	13	75
December 2011 (autumn)	6-17	18-24	10-12	7.5	345	520	183.8	42.5	32.8	17.3	39.0	0.05	0.2	0.04	1.0	472	327	14	17

Table (3): Effect of average of physico-chemical and seasonal parameters on the number of fungal collections.

NRS = number of fungal collections per sample; As R = ascomycetes records, An R=anamorphic fungi record; BR = Basidiomycetes records; MR= Myxomycetes records.

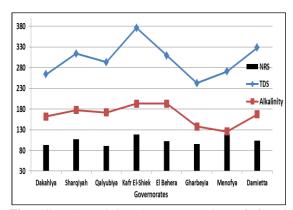


Fig. (1): The relation between number of fungal collections per sample and averages of TDS, alkalinity in the eight studied governorates (NRS \times 10²).

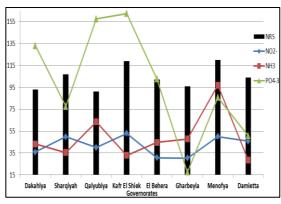


Fig. (2): The relation between number of fungal collections per sample and averages of NO₂, NH₃ and PO₄³⁻ ions in the studied governorates (NRS, NO₂, NH₃, PO₄³⁻x10²).

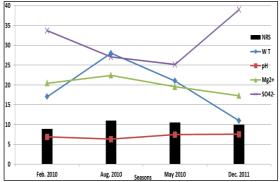


Fig. (3): The relation between number of fungal collections per sample and averages of water temperature, pH, Mg^{2+} , SO_4^{2-} in the studied seasons (NRS x 10).

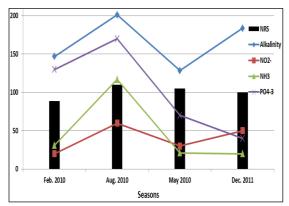


Fig. (4): The relation between number of fungal collections per sample and averages of alkalinity, NO_2 , NH_3 and PO_4 ³⁻ ions in the studied seasons (NRS, NO_2 , NH_3 , PO_4 ³⁻ x 10²).

The relationship between fungal distribution parameters and physichochemical analysis:

By using direct observation to the results, we cannot put constant base or direct between physicho-chemical analysis and biodiversity of fungi for example: the highest number of collection per sample was recorded in Menofya governorate in August collection and also the highest average of NO₂⁻ and NH₃⁺ were recorded in the same governorate. Otherwise, approximately the lowest average of TDS, alkalinity and Mg²⁺ were recorded in Menofya governorate (Tables 2 and 3). The lowest number of collection per sample in December 2011 was reported in Sharqiyah governorates, where concentration of Ca⁺² and SO₄²⁻ was high. Many other examples of correlation patterns between physico-chemical parameters and fungal population were in Tables (1,2 and 3) and Figs. (1 to 4). In this study ascomycetes and anamorphic fungi records were high in governorates with low salinity (e.g. Dakahlya, Qalyubiya and Gharbeyia governorates). Some family like Halosphaeriaceae exhibited more preference to governorates with high total dissolved salts; Damietta, Sharqiyah and KafrEl-sheikh governorates.

We try to conclude clear relation between physico-chemical results and biodiversity of fungi using correspondence analysis (CCA). The environmental data were correlated with frequency of occurrence of fungal species data set, and the results for the CCA of governorates in relation to environmental variables were given in Table (4) and Fig. (5). Canonical correspondence analysis (CCA) triplot explained 41.6% of the variability in the fungal community and environmental data with 22.2% in axis 1 and 19.4% in axis 2 (Table 4). The orientation environmental axis reflects the direction of maximum change of that variable. The longer of the axis indicate to the strength of influence of variable on community composition. Thus sites near or beyond the tip of an axis will be strongly positively correlated with and influenced by the environmental variable represented by that arrow. Axis 1 negatively correlated with PO₄³- and EC variables Likewise, a significant positive correlation of axis 1 was observed with pH average values. A significant positive correlation of axis 2 with NH₃ was observed, while it was significantly negatively correlated with alkalinity, TDS, Ca+2 and Mg2+ (Table 4 and Fig 5). There was no significant correlation of SO₄² and NO₂ with both axes.

	CCA Axes							
Variables	1	2						
Eigenvalue (%)	0.237 (22.2%)	0.207 (19.4%)						
pН	0.46	0.12						
TDS	0.01	-0.77						
EC	-0.33	-0.45						
Alkalinity	-0.08	-0.86						
Cl-	-0.29	0.50						
Ca ⁺²	-0.03	-0.60						
Mg^{2+}	-0.13	-0.58						
SO4 ²⁻	0.11	0.14						
NO ₂ -	0.11	-0.07						
NH3	-0.30	0.87						
PO ₄ ³ -	-0.42	-0.20						

Table 4: Summary of canonical correspondence analysis (CCA) using PAST software.

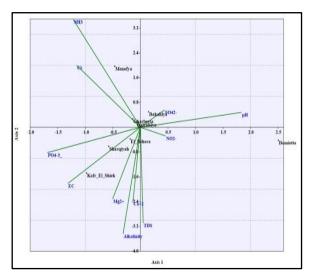


Fig. 5: Canonical correspondence analysis (CCA) plot explaining impact of environmental variables on species distribution

Discussion:

This part of the present investigation was planned to monitor the physico-chemical properties of water samples in river Nile streams in Delta region and study the effects of the changes in fungal distribution in the different governorates and seasons. The River Nile and its streams in Egypt can be characterized to high, moderately and low polluted. The Nile water is of high quality as the river reaches Cairo. Deterioration in water quality occurs when the Nile splits into the Damietta and Rosetta branches in a northward direction due to industrial effluents and agricultural drainage with decreasing flows (World Bank 2005).

The distribution pattern of fungi within streams can be profoundly altered by human activities, including the input of nutrients, sewage, pesticides and industrial wastes (Bärlocher 1992b). Factors affecting colonization of fungi involve types of substratum, temperature, relative humidity, the season of sampling, water chemistry and geographical location (Kohlmeyer and Kholmeyer 1979; Hudson 1986). Air and water temperature were ones of the main

factors effecting on the fungal distribution between different seasons collections. The highest species richness and ascomycete records were observed in summer and May collections while anamorphic fungi records were high in February and December collections. Shearer (1972) and Suberkropp (1984) found that seasonal changes in temperature affected the occurrence of aquatic ascomycetes and hyphomycetes.

Chamier (1992) reviewed a number of important factors that effect on the fungal communities inside the stream such as water temperature, conductivity, pH, nitrate and phosphorus concentration. Furthermore, Hu et al. (2010) suggested that changes in water aeration, pH and turbidity may be causal factors on fungal communities. In this investigation pH values was slightly fluctuated and not exceeded 7.8 and contrary correlated with most fungal distribution parameters in Nile Delta region. The pH value is particularly play a crucial role in determining the fungal diversity and their composition. Dubey et al. (1994) stated that the increase in water acidity lead to reduce the diversity of aquatic hyphomycetes in West Virginia. Also, Bärlocher and Rosset (1981) reported that the fungal diversity in soft water streams was higher than those in hard water streams where the pH was higher. The fluctuation in pH arises from different industrial effluents and agricultural drainage in waters.

Electric conductivity of Nile water samples exhibited great differences between the different governorates and different season's samples. Electric conductivity and alkalinity have a contrary effect on the most fungal distribution parameters in studied governorates and seasons. The increase in the electric conductivity was resulting from agricultural drainage which has high conductivity (APHA 1998; Gali *et al.* 2012). Salinity played active role in determining fungal diversity and community composition in this study. Less is known about the variation of fungal diversity

and composition along latitude, altitude and salinity gradients (Mohamed and Martiny 2010). In general, there was decrease in fungal diversity with increasing salinity in this study.

Ammonia and phosphate have enhancing effect in most distribution parameter in this investigation. The level of nitrates and phosphate in water has been regarded as important limiting factors in determining rates of detritus degradation (Elwood et al. 1981; Suberkropp and Chauvet 1995). Sridhar and Bärlocher (1997) found that the sporulation of aquatic hyphomycetes was stimulated by increasing nitrates. The presence or increase in NO₂ and NH₃ in some streams may be resulting from fertilizer that may be present in the soil and it easily oxidized to nitrite (Karavoltsos et al. 2008). There was no significant correlation of SO_4^{2-} and NO_2^{-} with fungi reported during this study. Suberkropp et (1988)found that the aquatic hyphomycetes community was unaffected by the effluents discharged from a sewage treatment plant.

Shearer (1972) studied the distribution of aquatic ascomycetes at 5 stations along river in Chesapeake Bay. She found that 6 species collected at station 1 and 3 were not collected at station 2 and suggested that was probably because of the effluents discharged from a sewage treatment plant near station 2. Tsui et al. (2001) investigated the longitudinal and distribution temporal of freshwater ascomycetes and hyphomycetes on submerged wood in the Lam Tsuen River in Hong Kong and found that the fungal species composition changed correlated with concentrations of NO3 -N, NH₃ -N, and PO₄ -P resulting from human disturbance. Luo et al. (2004) compared the fungal communities and diversity between Lake Fuxian (an unpolluted lake) and Lake Dianchi (a heavily polluted lake), and concluded that pollution causes change in the fungal communities, but had little effect on fungal diversity. The number of fungal species in submerged wood may not be good bio- indicators of organic pollution (Tsui et al. 2001). Au et al. (1992) and Raviraja et al. (1998) compared the species diversity in unpolluted and polluted streams located geographically close together. They observed that freshwater fungi diversity, species richness and conidial production were reduced with organic pollution in the polluted streams.

 Ca^{+2} and Mg^{2+} concentration negatively correlated with fungal communities. However, physico-chemical parameters such as SO₄²⁻ and NO₂⁻ they were not having significant correlation on fungal communities. Many metals e.g., copper (Cu), magnesium (Mg), potassium (K), sodium (Na), iron (Fe) are essential to fungal growth, development and differentiation (Hughes and Poole 1991). However, metals can be toxic above a critical concentration that depends on the organism, the physico-chemical properties of the metal, and the environmental factors (Gadd 1993; Blaudez et al. 2000; Gimmler et al. 2001). Although fungal diversity has been recognized to decrease due to heavy metal exposure (Bermingham et al. 1996), several species of aquatic hyphomycetes have been found in metal-polluted streams (Krauss et al. 2001; Pascoal et al. 2005).

Fluctuation observed here in physichochemical parameters are probably due to combination of several factors including industrial, agricultural and domestic waste discharge. Results from CCA analysis of the fungal assemblages found at the eight governorates revealed that changes in species composition were correlated with elevated concentrations of NH₃, and pH resulting from domestic and agricultural activities in River Nile streams.

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الملخص العربي

العلاقه بين الخواص الكيميائيه والفيزيائيه لتفرعات نهر النيل والتنوع الفطري في منطقه دلتا مصر

تلوث مياه نهر النيل في مصر في ازدياد دائم مع استمرار تدفق النفايات بشكل مستمر. عينات من المياه جمعت من ترع وتفرعات نهر النيل الاساسيه من ثمان محافظات في الدلتا علي مدار اربع مواسم مختلفه في الفتره من فبراير 2010 الي ديسمبر 2011. تم دراسة العديد من العلاقات بين خواص المياه الكيميائيه والفيزيائيه والتنوع الفطري في منطقة الدلتا. درجة حرارة الهواء والماء كانت من اكثر العوامل الاساسيه الموثره علي توزيع الفطريات. تركيز الكالسيوم والمغنسيوم كان له تاثير عكسي علي تواجد الفطريات، بينما تواجد الكبريتات والنيتريت لم يكن له تأثير واضح. الأمونيا والفوسفات كان لهما تأثير محفز لتوزيع الفطريات في منطقة الدلتا. ملوحة المياه كان لها تأثير واضح علي تحديد نوعية وتنوع الفطريات في منطقة الدراسه.